

THYRSIFEROL: A SQUALENE-DERIVED METABOLITE OF LAURENCIA THYRSIFERA

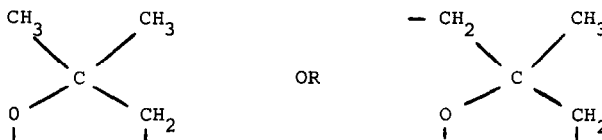
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Previous investigations of red algae of the genus Laurencia (Rhodomelaceae, Rhodophyta) have established the structures of many interesting metabolites based in the main on fatty acid, sesquiterpenoid or diterpenoid skeletons.<sup>1</sup> We now report the isolation and identification of a squalene-derived metabolite from a Laurencia species.

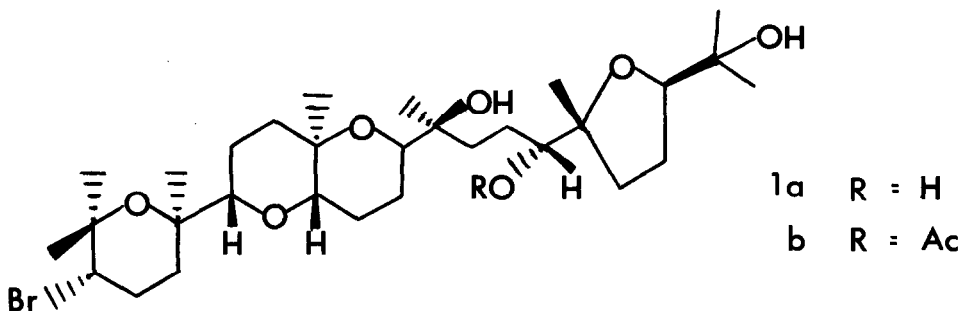
Laurencia thyrsoifera (Hook) was collected from the intertidal zone at Seal Reef, Kaikoura, N.Z.<sup>2</sup> Chromatography of the methanol extract (4.5% of dry weight) on alumina gave a complex mixture of fatty acid methyl esters,<sup>3</sup> cholesterol<sup>4</sup> and the polar compound thyrsoiferol (1a) as a mixture with two other metabolites (ene-yne)s. After acetylation<sup>5</sup> of this mixture thyrsoiferol (1a) was isolated as its mono-acetate (1b), m.p. 104.5-106°, RD (in cyclohexane, c = .0002)  $[\alpha]_{400} + 89^\circ$ ,  $[\alpha]_{350} + 111^\circ$ ,  $[\alpha]_{300} 220^\circ$ ,  $[\alpha]_{225} 400^\circ$ . The molecular formula was inferred from high resolution mass measurement of the  $[M-H_2O]$  doublet and the  $[M-HBr]$  peak, as the molecular ion doublet (m/e 646/648) was of low intensity. The <sup>13</sup>C nmr spectrum<sup>7</sup> confirmed that the acetate (1b) was highly oxygenated as eleven C-O type resonances ( $\delta_{13C}$  60-80) were discernible. The molecular formula, <sup>13</sup>C offset resonance decoupling experiments and <sup>13</sup>C LIS studies established that thyrsoiferyl acetate (1b) was a saturated, non-carbocyclic, dihydroxy acetate with 4 ether bridges. In addition, some 8 methyl groups were resolved (excluding the acetate methyl) and these were in similar structural environments:



The structure of thyrsoiferyl acetate (1b) was determined by X-ray crystallography. Preliminary photographs indicated that thyrsoiferyl acetate belonged to the monoclinic crystal class and systematic absences ( $OkO$  with  $k$  odd) together with the chirality of the compound suggested that the space group was  $P2_1$ . Refined cell constants were  $a = 12.348(4) \text{ \AA}$ ,  $b = 12.055(5) \text{ \AA}$ ,  $c = 12.166(5) \text{ \AA}$  and  $\beta = 107.31(4)^\circ$ ,  $D_c = 1.24 \text{ g cm}^{-3}$  and  $Z = 2$ .<sup>8</sup> The best crystal available for intensity collections was of high mosaicity ( $0.36-0.48^\circ$ ) with dimensions

of  $(0.250 \times 0.075 \times 0.063)\text{mm}^3$ . A total of 1931 unique reflections ( $\theta < 50^\circ$ ) were collected using Ni-filtered Cu K $\alpha$  radiation ( $\lambda = 1.5418 \text{ \AA}$ ) with  $\theta$ - $2\theta$  scans on a Hilger and Watts four-circle automatic diffractometer. Of these reflections 1103, for which  $F_o^2 > 3\sigma(F_o^2)$ , were used in all structure solving calculations. Lorentz and polarisation corrections were applied, but no absorption corrections were made ( $\mu = 21.53 \text{ cm}^{-1}$ ).

The crystal structure was determined by conventional heavy atom methods and refined using the full-matrix least-squares procedure with isotropic temperature factors for all carbon and oxygen atoms and anisotropic temperature factors for the bromine atom.<sup>9</sup> The scattering factors of Cromer and Mann<sup>10</sup> with the anomalous dispersion corrections of Cromer<sup>11</sup> for bromine were applied. Hydrogen atoms were not included in the structure factor calculations. This model was refined to conventional agreement factors  $R = 0.144$  and  $R_w = 0.182$ . The centrosymmetric distribution of bromine atoms in an otherwise non-centrosymmetric array, together with the poor crystal quality, severely hindered attempts to determine the absolute configuration. The correct enantiomer has not been conclusively established but the weighted R-factor and the standard error in an observation of unit weight were marginally better for the enantiomer shown. Final position and thermal parameters are given in Table 1.<sup>12</sup>



The conformation of thyransferyl acetate in the crystal studied is shown in Fig. 1. Ring C is in a twist-boat conformation in which strong 1,3-diaxial interactions between C(27) and C(15) are avoided. The molecular packing is dominated by intermolecular H-bonding (2.80(4)  $\text{\AA}$ ) between O(4) and O(6) in adjacent molecules, and there is close intra-molecular contact between O(7) and O(6) (2.82(4)  $\text{\AA}$ ).

Limited biological testing<sup>13</sup> of a crude sample of thyransferol (1a) and thyransferyl acetate (1b) indicated that they were biologically inactive.

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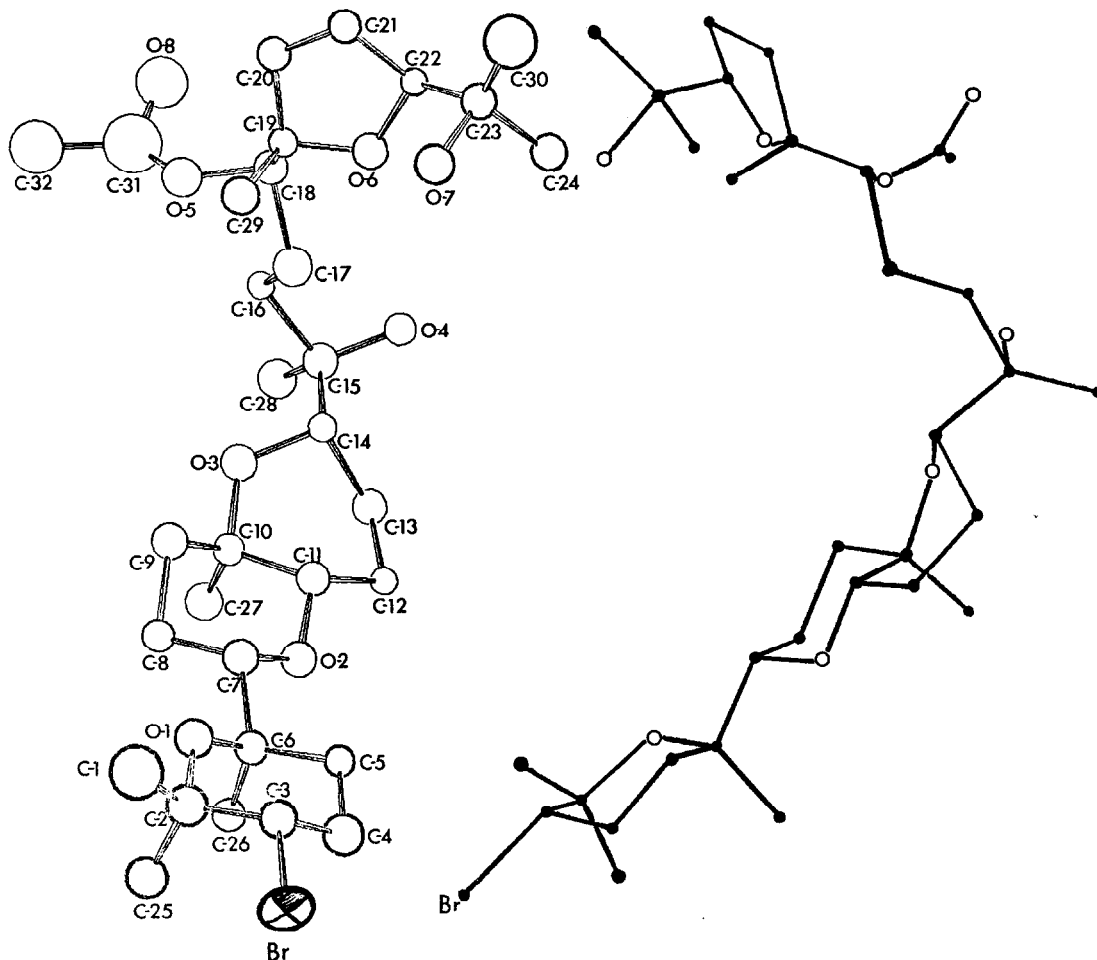


Fig. 1: Two perspective views of thyransferyl acetate (1b).

	x	y	z	U		x	y	z	U
Br	0.2389(4)	0.2500	0.0263(5)		C(13)	0.672(4)	1.014(4)	0.265(4)	0.06(1)
O(1)	0.517(2)	0.438(2)	0.232(2)	0.060(9)	C(14)	0.796(3)	0.958(4)	0.252(3)	0.04(1)
O(2)	0.540(2)	0.734(3)	0.249(2)	0.057(9)	C(15)	0.879(4)	1.362(4)	0.300(4)	0.06(1)
O(3)	0.818(2)	0.874(3)	0.346(2)	0.061(9)	C(16)	1.007(3)	0.996(3)	0.302(3)	0.03(1)
O(4)	0.855(2)	1.133(2)	0.204(2)	0.049(8)	C(17)	1.000(4)	0.944(4)	0.195(4)	0.07(1)
O(5)	1.185(3)	0.879(3)	0.274(3)	0.07(1)	C(18)	1.129(4)	0.943(4)	0.087(3)	0.05(1)
O(6)	1.067(2)	0.934(2)	-0.014(2)	0.056(9)	C(19)	1.143(3)	0.884(3)	0.171(4)	0.04(1)
O(7)	0.991(2)	0.807(3)	-0.218(2)	0.063(9)	C(20)	1.249(3)	0.886(4)	0.052(3)	0.04(1)
O(8)	1.314(4)	1.000(4)	0.349(4)	0.12(1)	C(21)	1.223(3)	0.891(4)	-0.079(3)	0.03(1)
C(1)	0.511(5)	0.240(7)	0.186(5)	0.11(2)	C(22)	1.109(3)	0.949(3)	-0.116(3)	0.05(1)
C(2)	0.441(4)	0.334(4)	0.199(4)	0.06(1)	C(23)	1.033(4)	0.922(4)	-0.236(4)	0.05(1)
C(3)	0.354(3)	0.365(4)	0.080(4)	0.06(1)	C(24)	1.073(5)	0.902(6)	-0.339(5)	0.11(2)
C(4)	0.292(4)	0.484(4)	0.075(4)	0.07(1)	C(25)	0.389(4)	0.312(4)	0.291(4)	0.07(1)
C(5)	0.383(3)	0.571(3)	0.108(3)	0.04(1)	C(26)	0.412(3)	0.572(4)	0.328(4)	0.05(1)
C(6)	0.467(3)	0.549(4)	0.232(4)	0.05(1)	C(27)	0.688(4)	0.831(4)	0.453(4)	0.06(1)
C(7)	0.582(4)	0.615(4)	0.249(4)	0.06(1)	C(28)	0.897(4)	1.112(4)	0.425(4)	0.07(1)
C(8)	0.671(3)	0.602(3)	0.363(4)	0.05(1)	C(29)	1.109(3)	0.741(5)	0.075(3)	0.06(1)
C(9)	0.774(4)	0.687(4)	0.357(4)	0.06(1)	C(30)	0.926(4)	0.999(4)	-0.260(4)	0.05(1)
C(10)	0.725(3)	0.803(4)	0.357(4)	0.04(1)	C(31)	1.264(7)	0.913(8)	0.364(7)	0.15(2)
C(11)	0.625(4)	0.809(4)	0.237(4)	0.05(1)	C(32)	1.332(5)	0.845(6)	0.468(6)	0.12(2)
C(12)	0.572(3)	0.928(3)	0.216(3)	0.04(1)					

	U <sub>11</sub>	U <sub>22</sub>	U <sub>33</sub>	U <sub>12</sub>	U <sub>13</sub>	U <sub>23</sub>
Br	0.069(3)	0.044(3)	0.119(5)	-0.040(3)	0.013(3)	-0.034(4)

Table 1: Position and thermal parameters for thyransferyl acetate (1b)

References and Notes:

1. D. J. Faulkner, Tetrahedron, **33**, 1421 (1977), W. Fenical, J. Phycol, **11**, 245 (1975) and references cited therein.
2. Collected at low tide on 2 February, 1975.
3. Identified by glc retention time on DEGS and by gc/ms: 12:0, 14:0, 16:0, 16:107, 18:0, 18:109, 18:1010, 18:206, 18:203, 18:303, 18:403, 20:309, 20:306, 20:406.
4. Identified on the basis of mp, mixed mp,  $^1\text{H}$  nmr,  $^{13}\text{C}$  nmr, ms.
5. Pyridine/acetic anhydride (1:1).
6. AEI MS-902C, Varian-MAT CH-5.
7. Varian CFT-20. Spectrum acquired in  $\text{CDCl}_3$ , referenced to TMS at  $\delta_{^{13}\text{C}}$  0.
8. Standard deviations for the last significant digit are given in parentheses.
9. Calculations were carried out on a Burroughs B6718 computer using local modifications of the well-known programs ORFLS and ORFFE (W. R. Busing, K. O. Martin and H. A. Levy) and FORDAP (A. Zalkin).
10. D. T. Cromer and J. B. Mann, Acta Cryst., **A-29**, 321, (1968).
11. D. T. Cromer, Acta Cryst., **18**, 17 (1965).
12. Isotropic thermal parameters in  $\text{\AA}^2$ . Anisotropic parameters (U's), also in  $\text{\AA}^2$ , are coefficients in:  

$$\exp -2\pi^2 (U_{11}h^2a^{*2} + U_{22}k^2b^{*2} + U_{33}l^2c^{*2} + 2U_{12}hka^*b^* + 2U_{13}hla^*c^* + 2U_{13}hla^*b^* + 2U_{23}klb^*c^*)$$
13. Test organisms used were: Bacillus subtilis, Staphylococcus aureus, Pseudomonas aeruginosa, Aspergillus niger, Saccharomyces cerevisiae.